

The association between ABO blood groups and norovirus infections among patients suffering from diarrheal disease in Northwest Ethiopia

Dessie Tegegne^{1,2*}, Aschalew Gelaw¹, Demeke Endalamaw³, Getachew Ferede¹, Baye Gelaw¹

¹Department of Medical Microbiology, School of Biomedical and Laboratory Sciences, College of Medicine and Health Sciences, University of Gondar, Gondar, Ethiopia, ²Department of Medical Laboratory Sciences, College of Health Sciences, Debre Tabor University, Debre Tabor, Ethiopia, ³Department of Medical Microbiology and Virology, Amhara Public Health Institute, Bahir Dar, Ethiopia

*Corresponding author: Email: desstegegne100@gmail.com

Citation: Tegegne D, Gelaw A, Endalamaw D, Ferede G, Gelaw B. The association between ABO blood groups and norovirus infections among patients suffering from diarrheal disease in the Amhara National Regional State, Ethiopia: ABO blood group and norovirus infection. *Ethiop J Health Biomed Sci.* 2024;14(1):45-53.

DOI: <https://doi.org/10.20372/ejhbs.v14i1.707>

Article History

Received: March 18, 2024

Revised: May 20, 2024

Accepted: May 28, 2024

Key words: blood groups, norovirus, diarrheal patients, all ages

Publisher: University of Gondar

Abstract

Background: Norovirus (NoV) infection is a significant cause of diarrhea worldwide. However, all individuals are not equally affected due to environmental, viral, and host factors, particularly ABO blood groups. Indeed, data that describes the association between NoV infection and the ABO blood group is limited in Ethiopia, and this needs to be investigated.

Objective: This study aimed to assess the relationship between ABO blood groups and NoV infection in Northwest Ethiopia.

Method: A health-facility-based cross-sectional study was conducted from May 2021 to November 2021 by enrolling 550 participants with diarrhea. Fecal samples were collected and analyzed by reverse transcription PCR to identify NoVs. To further genotype the positive samples, a viral protein-1-coding gene was sequenced. In addition, blood samples were collected and tested to identify blood groups by using the tube hemagglutination technique. The data were analyzed using SPSS version 23. Logistic regression analysis was done to assess the association between NoV infection and the independent variables.

Result: Among the 550 enrolled participants, 519 (94% response rate) provided the required clinical samples and epidemiological data. The majority of the study participants (249/519; 48%) had O blood group. Among the NoV-positive study subjects, the majority (34/46, 74%) were in blood group O, followed by blood group A individuals (9/46, 19.6%). The risk of NoV infection was higher for patients with blood group O than for blood group B (AOR = 1.5, 95%CI = 2–15, P = 0.01), but there was no association for other blood groups. At least one NoV-GII was identified in each of the blood groups, while NoV-GI affected individuals with blood groups O and A. Besides, GII.3 and GII.21 genotypes were common among blood group O individuals, while most (75%) blood group A individuals were susceptible to GII.17 infections.

Conclusion: The positivity rate of NoV infection was considerably high among individuals with blood group O. Norovirus-GII can infect all blood groups, while NoV-GI selectively affects blood groups A and O. Further large-scale studies are warranted to assess the relevance of this observation and other genetic factors.

Copyright: © 2022 at Tegegne et al. This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 (CC BY NC 4.0) License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Introduction

Diarrheal diseases are among the most common causes of morbidity and mortality, responsible for more than 1.5 million deaths globally (1). In developing countries, particularly, diarrhea is the second most common cause of death (2), with the highest burden resting in sub-Saharan Africa and south Asia (3). As a sub-Saharan African country, diarrheal disease is also an important public health concern in Ethiopia (4). More than 25% of under-5 children are affected by diarrhea (5, 6). Viruses are the leading etiological agents of diarrhea in both high- and low-income countries (7). Norovirus (NoV) infections pose a significant burden of diarrheal diseases among all age groups globally, with variable distributions across different settings (8, 9). These viruses are responsible for about 20% of all episodes of diarrheal disease globally (1). The estimated burden is 699 million cases and 219,000 deaths annually, where 97% of mortality is attributed to lower and middle-income countries (1, 10).

Human NoVs are genetically diverse groups of viruses under the family *Caliciviridae*. The virion contains a non-enveloped single-stranded (+sense) RNA genome (11). Based on the partial viral protein 1 (VP1) sequence analysis, they are classified into 10 genogroups (GI to GII) and 49 genotypes. Of these genogroups, GI, GII, and GIV are known to infect humans. These NoV genogroups are further divided into multiple genotypes (9 GI, 27 GII, and 2 GIV) (12). Norovirus GII is by far the predominant genogroup (13), and NoV-GII, genotype-4 (GII.4), is reported as the predominant genotype globally (9, 14). However, a predominance of non-GII.4 genotypes (GII.3, GII.17, and GII.2) over GII.4 was reported in the past five years (15, 16). This might be related to their rapid evolution and a change in their binding profiles (17).

Noroviruses are highly contagious, with an estimated infectious dose of as few as 18 viral particles and prolonged shedding over an average of 8–60 days (18, 19). The main route of transmission is direct contact with an infected person, and it can also be transmitted via contaminated food and water (20, 21). Despite the high infectivity and rapid transmission, not all individuals are affected and develop symptomatic disease (22).

The association between ABO blood groups and NoV infection was reported previously, but with a conflicting result (23, 24).

One previous study demonstrated that NoV recombinant virus-like particles strongly agglutinated red blood cells from group O, A, and AB donors but were less likely to do so from group B individuals (25). Another study also reported that among NoV-positive individuals, all symptomatic infections were among blood group O and group A individuals (26). In another study, genogroup-specific susceptibility was reported with NoV-GI, which mainly targets individuals with blood groups O and A (27), while NoV-GII infected all individuals irrespective of their ABO blood group status (28). In contrast, a partial or total absence of correlation between different blood group (A, B, AB, and O) antigens and NoV infection was reported (29–31).

The distribution of ABO blood groups and other host genetic factors is strongly dependent on ethnicity (32, 33). Besides, the molecular epidemiology of NoV differs between different areas or countries due to different environmental, viral (34–36), and host factors (22, 37, 38). However, studies addressing the association of host factors, especially ABO blood groups, with NoV infection are limited in Africa and have not yet been conducted in Ethiopia. Therefore, this study aimed to assess the relationship between ABO blood groups and NoV infection in Northwest Ethiopia.

Method

Study design, study period, and setting: From May 2021 to November 2021, a health-facility-based cross-sectional study was conducted to collect clinical and epidemiological data. Four data collection sites (Debre Markos, Bahir Dar, Gondar, and Debre Tabor), all of which are found in Northwest Ethiopia, were considered. In each of the study sites, one comprehensive specialized hospital and two health centers were included. The health institutions included in Debre Tabor were Debre Tabor Comprehensive Specialized Hospital, Liul Alemayehu Health Center, and Debre Tabor Health Center. In the second study site, Bahir Dar, Felege Hiwot Comprehensive Specialized Hospital, Shimbit Health Center, and Abay Health Center were included. Similarly, Gondar consists of the University of Gondar Comprehensive Specialized Hospital, Azezo Health Center, and Gondar Health Center. Lastly, in Debre Markos, Debre Markos Comprehensive Specialized Hospital, Debre Markos Health Center, and Gozamen Health Center were considered.

Sample size determination and sampling technique: A single population proportion formula was used to calculate the sample size ($n = Z_{\alpha/2}^2 * P(1-P)/d^2$). By considering the following assumptions: $Z_{\alpha/2}$ is taken as 1.96 at a 95% confidence interval (CI); P is the proportion taken from the previous study (13.3%)(39); d is the desired level of precision (3%). Finally, a 10% non-response rate was added, and the total sample size was calculated at 550. Based on the previous year's diarrheal disease case flow, the total sample size was proportionally allocated as 100, 120, 152, and 178 for each of the Debre Tabor, Debre Markos, Gondar, and Bahir Dar sites, respectively. A systematic random sampling technique was used to select the study participants from each health facility.

Data collection: Socio-demographic data were collected by trained healthcare professionals working in each of the health facilities. Once a clinical diagnosis was made based on the inclusion criteria, a pre-tested, semi-structured questionnaire was administered to each participant.

Fecal sample collection, processing, and molecular characterization: Five mL diarrheic fecal samples were collected using sterile containers from individuals of all age groups with diarrhea and self-reported to the health facilities during the study period. These fecal samples were stored at $-20\text{ }^{\circ}\text{C}$ or lower in each of the health facilities until transported to the Amhara Public Health Institute (APHI) for molecular investigation. Besides, peripheral blood samples were also collected to assess the blood group status of study participants. Once the fecal samples were taken to the APHI, they were stored at $-70\text{ }^{\circ}\text{C}$ until processed. There, ten percent (weight/volume) of fecal suspensions were prepared with nuclease-free water or 1% phosphate-buffered saline as available, vortexed, and centrifuged. Viral ribonucleic acid (RNA) was extracted from 300 μL fecal suspensions using MagaBio Plus Virus RNA Purification Kit II (Hangzhou, China). The details for the detection and sequencing or genotyping protocols, list of primers and probes that used for screening and genotyping as well as all PCR conditions are available in our recent publication (40).

Blood sample processing and ABO blood grouping: The assigned and trained health care worker in each ward of the health facilities informed the patient that he had received permission to collect epidemiological data and sent him with a request paper to a laboratory room for blood group analysis.

The trained laboratory professional, in the laboratory room, instructs the client and collects 5 mL of blood. Then blood typing was done on-site at each of the study sites with a tube hemagglutination technique as previously done(41). Briefly, a forward grouping protocol was applied to assess the presence or absence of A, B, and D antigens in the red blood cells using commercially prepared antisera (Cypress Diagnostics, Langdorp, Belgium). Blood cells were placed in the three test tubes to prepare a 5% suspension of red blood cells to be tested in isotonic saline. Then one drop of each RBC suspension in each tube was mixed with a drop of anti-A, anti-B, and anti-D. These tubes were subjected to centrifugation for 3 minutes to ensure enhanced chemical interactions, particularly for weaker antibodies to react and agglutinate. The resultant matrix was gently shaken and examined macroscopically for agglutination. The tubes were categorized according to the extent of blood clumping as A+, A-, B+, B-, AB+, AB-, O+, and O-.

Statistical analysis: Data were entered and analyzed using SPSS version 23 software. Descriptive statistics were used for frequencies and percentages. The association between the outcome variables and factors was assessed using a logistic regression analysis. Those variables with P values < 0.05 and an adjusted odds ratio (AOR) in a 95% CI were considered statistically significant. The assumption is that the AOR within the 95% CI should not include 1. The result was interpreted and presented in a summary or displayed by using tables and figures.

Result

Socio-demographic characteristics: Among the total 550 study participants enrolled, 519 (with a 94.4% response rate) provided the required clinical samples and socio-demographic information. The age of the study participants ranged from 3 months to 85 years. More than half (51.3%) of the participants were female. From the four study sites, one-third (178/519; 34.3%) of the study participants were recruited from Bahir Dar, followed by Gondar (142/519; 27.4%). Three-fourths of the study participants were came from urban areas. In addition to this, about two-thirds (330/519; 63.6%) of our study participants and/or their parents were married. Besides, the majority (407/519; 78.4%) of the study participants were literate (they were at least able to read and write).

ABO blood groups and Rh types: We identified four blood groups (A, B, AB, and O) and assessed the Rh factor status. Nearly half (249/519; 48%) of the study participants had the O blood group, followed by blood groups A (143/519; 27.5%)

and B (103/519; 19.8%). More specifically, the majority (216/519; 41.6%) were identified as blood group O with an Rh factor (O+), which was followed by A+ (119/519; 22.9%) and B+ (88/519; 17%) (**Fig. 1**).

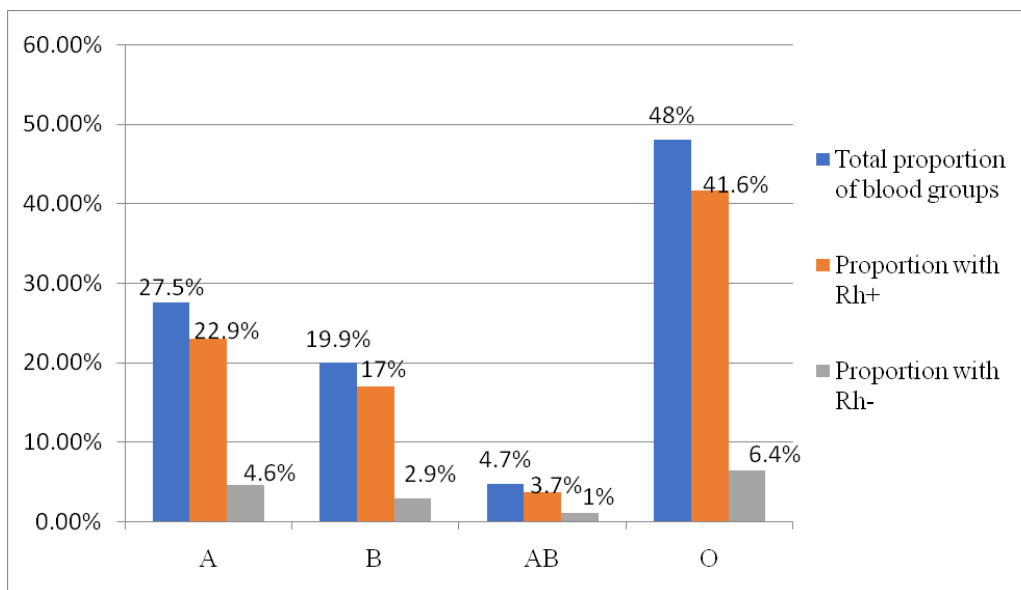


Figure 1: The proportion of ABO blood groups and Rh status among patients with diarrhea in Northwest Ethiopia; May 2021 to November 2021

Association between ABO blood groups and norovirus infection: Norovirus was identified in 8.9% (46/519) of the fecal samples of individuals with diarrhea. Both NoV-GI and GII were identified, with GII being predominant (38/46; 82.6%). The identified NoV genotypes were GI.3, GI.5, GII.3, GII.6, GII.10, GII.17, and GII.21. Of all the 29 successfully genotyped NoVs, genotype GII.3 was the predomi-

nant (13/29; 44.8%), followed by GII.21 (6/29; 20.7%) and GII.17 (4/29; 13.8%). In addition to this, GII.21 was identified for the first time in Ethiopia. Study participants with blood group O had the highest proportion of NoV infection (34/249; 13.7%), followed by blood group A (9/143; 6.3%), AB (1/24; 4.2%), and B 1/46 (2/103; 1.9%)(**Table 1**).

Table 1: The distribution of noroviruses across ABO blood groups of study participants in Northwest Ethiopia; from May 2021 to November 2021

NoV positivity, genogroups, and genotypes		ABO blood groups (%)			
		O	A	B	AB
Status of NoV infection	Positive	34 (13.7)	9 (6.3)	2 (1.9)	1 (4.2)
	Negative	215 (86.3)	134 (93.7)	101 (98)	23(95.8)
Genogroups	GI	6 (75)	2 (25)	0	0
	GII	28 (73.7)	7 (18.4)	2 (5.3)	1 (2.6)
Genotypes	GI.3	0	1	0	0
	GI.5	3	0	0	0
	GII.3	9	2	1	1
	GII.6	1	0	0	0
	GII.10	1	0	0	0
	GII.17	1	3	0	0
	GII.21	4	1	1	0

Among the NoV-positive study subjects, the majority (34/46, 74%) were blood group O, followed by blood group A individuals (9/46, 19.6%). Norovirus-GII was detected across participants with O, A, B, and AB blood groups, with the highest proportion (28/38; 73.7%) among blood group O participants. Based on the multiple logistic regressions analysis, the probability of NoV infection was increased among under-5 children (AOR = 1.4, 95% CI: 2.7–18, $P = 0.02$), the elderly (AOR = 5, 95% CI: 1.7–16, $P = 0.015$), individuals living in the Bahir Dar area (AOR = 1.5, 95% CI: 1.6–22, $P = 0.014$), and Debre Tabor (AOR = 2.5, 95% CI: 1.8–23, $P = 0.001$). Besides, the odds of

NoV infection among blood group O individuals were 1.5 times higher than B blood groups (AOR: 1.5, 95%CI: 2–15, $P = 0.001$). Norovirus-GI was detected only in participants with blood groups O and A. Except for GI.3, all the other NoV genotypes were detected in participants with blood group O. The GII.3 genotype was detected across the four blood groups. Similarly, GII.21 was identified in participants with blood groups A, B, and O. In addition to this, three-fourths (3/4; 75%) of the GII.17 genotypes were detected in participants with blood group A. However, the difference was not statistically significant (P value > 0.05)(Table 2).

Table 2: The association of ABO blood groups and other variables with norovirus infection among patients with diarrhea in Northwest Ethiopia; May 2021 to November 2021

Variable	Categories	Norovirus status		COR (95% CI)	Pvalue	AOR (95% CI)	Pvalue
		Positive	Negative				
		N (%)	N (%)				
Sex	Male	25 (10)	228 (90)	1.3 (0.7-2.3)	0.4		
	Female	21(8)	245 (92)	1			
Age group in years	<5	20 (12.5)	160 (87.5)	5.6 (1.9-16.7)	0.002	1.4 (2.7-18)	*0.02
	5 to 17	9 (8.2)	110 (91.8)	3.5 (1.3-9)	0.011	1.1 (0.2-17)	0.11
	18 - 64	9 (4)	225 (96)	1	1		
	> 64	8 (33.3)	24 (66.7)	12 (4-35)	0.000	5 (1.7-16)	*0.015
Blood groups	O	34 (13.7)	215 (86.3)	2.7 (3.6-12)	0.02	1.5 (2-15)	*0.01
	A	9 (6.3)	134 (93.7)	0.9 (0.08-5.3)	0.6	0.7 (0.5-8)	0.25
	AB	1 (4.2)	23 (95.8)	0.8 (0.2-25)	0.8	0.4 (0.2-14)	0.7
	B	2 (1.9)	101 (98)	1		1	
Study area	Debre Tabor	17 (17.2)	82 (82.8)	5 (1.6-15.4)	0.005	2.5 (1.8-23)	*0.001
	Bahir Dar	15 (8.4)	163 (31.4)	2.7 (1.2-6.25)	0.017	1.5 (1.6-22)	*0.014
	Gondar	10 (7)	132 (25.4)	2.25 (1.01-4.7)	0.032	2.8 (0.9-20)	0.13
	Debre Markos	4 (4)	96 (18.5)	1		1	

AOR, Adjusted odds ratio; COR, Crude odds ratio; *statistically significant

Discussion

In the present study, we tried to demonstrate the association between the ABO blood group and NoV infections by analyzing blood and fecal samples collected from diarrheal patients recruited from the four major and proximate cities located in Northwest Ethiopia. In this study, NoV was predominantly identified in the participants with blood group O (13.7%) followed by blood group A (6.3%). Despite a significant number of participants, with blood group B involved, only about 2% of them were positive for NoV infection.

Three-fourths of the study participants who were positive for NoV infection had O blood group, while blood groups B and AB were less likely to be infected with NoV. Our findings are in agreement with different studies (24, 31, 42). Besides, in the present study, at least one NoV-GII was identified in each of the four blood groups, while infection with NoV-GI was observed among individuals with blood groups O and A. This is in agreement with a few previous studies (27, 31). Protection of individuals with blood groups B and AB for both genogroups was also reported previously, which supports our finding(22, 24).

Our findings also showed that at least one genotype of NoV-

GII was found among individuals with O blood groups. Besides, the predominant genotype (GII.3) was identified in all of the blood groups, more commonly among blood group O and blood group A individuals. This finding is in agreement with a study conducted in China(43). Although it was difficult to compare as it was a systematic review and meta-analysis, significant susceptibility patterns of O blood groups were also reported in China, which supported our finding (24). An absence of correlation among all blood groups (A, B, AB, and O) with NoV infection was also reported (29-31). This might be due to the existence of different factors other than the ABO blood group system that play a great role in the susceptibility of NoVs (11, 44).

In the present study, GII.21 was identified among all blood groups except AB blood groups. However, this justification requires further investigation. In addition to this, GII.17 genotypes were identified among individuals with blood groups A and O. In contrast to this, one previous study reported that one GII.17 infection was found in an individual with blood group O (31). The difference might be explained by host factors other than the ABO blood group, including the secretory antigen status of the individuals. Most individuals, termed secretors (having an active fucosyltransferase (FUT2) gene), express the histoblood group antigens in different body fluids that commonly act as receptors for NoV. Hence, they are commonly sensitive to some NoV infections, while others with inactivated or mutated FUT2 genes, termed non-secretors, are resistant to NoV(11). It might also be due to the host microbiota (like *Enterobacter cloacae* and *Escherichia coli*) that can mediate NoV attachment to the host via HBGA-like carbohydrates expressed on the surface of these bacteria(44). Moreover, the previous studies, the presence of Lewis^b antigen among secretors(45), and Lewis^a antigen among non-secretors (46) might play a role in NoV attachment.

In our study, there was a significant difference in the positivity rates of NoV across the different age groups, with the extreme age groups being significantly affected. Our finding is in agreement with some studies (47, 48). The possible explanation for the increased susceptibility of these age groups might be either due to their increased environmental exposure (49) or their weakened immune system (50). Similarly, our findings varied with study sites, where a significant increase was reported at the Debre Tabor and Bahir Dar sites. This might be explained either due to the increased diversity of the population and contact-mediated infection in Bahir Dar sites and/or due to the

variability of weather conditions (relatively lower temperature, increased humidity, and increased rainfall) in the case of Debre Tabor sites that favor this virus replication (51, 52).

This is the first study conducted to assess the association between ABO blood groups and NoV infection in Ethiopia by considering multiple sites across all age groups. However, other host genetic factors, including secretory antigens and Lewis antigens, that might have a relationship are not assessed.

Conclusion and recommendation

The positivity rate of NoV was significantly higher among individuals with O blood groups compared to others. Norovirus-GII infected individuals of all blood groups, while NoV-GI affected blood groups A and O. This study might provide an input for viral-host interaction studies and vaccine design programs for NoV. Further large-scale studies are warranted to assess the relevance of this observation and identify other genetic factors that might affect the susceptibility of individuals to the different NoV genotypes.

Abbreviations:

AOR— Adjusted odds ratio

APHI— Amhara Public Health Institute

FUT2— Fucosyltransferase 2

NoVs— Noroviruses

NoV-GII— Norovirus-genogroup II

NoV-GI— Norovirus-genogroup I

ORF— Open reading frame

PCR— Polymerase chain reaction

RNA— Ribonucleic acid

RT-PCR— Real-time reverse transcription polymerase chain reaction

Declarations:

Ethical approval and consent to participate: The project was approved by the University of Gondar institutional review board with reference number V/P/RCS/05//765/2021. Written informed consent/assent was obtained either directly from the participants and/or their parents for children. The study was conducted according to the rules and regulations of the Helsinki Declarations. To maintain confidentiality, the data was recorded with a special code. The study participants were free not to participate or withdraw at any time.

Consent for publication: All authors agree to publish this manuscript in this journal

Data availability: The datasets supporting the conclusions are included within the manuscript. However, upon a reasonable quest, the data are available from the corresponding author [DT].

Competing interests: The authors declare that they have no conflict of interest.

Funding: This research was partially supported by the Debre Tabor University community service strengthening program award (budget code: DTU/RCC/6223) and the University of Gondar Post-Graduate Strengthening Program (budget code: UoG/PGRS/6223).

Authors contributions: DT, BG, AG, and GF all participated in the design and proposal development. DT and DE conducted the laboratory analysis. DT analyzed the data and drafted the manuscript. All the authors (DT, BG, AG, GF, and DE) revised the manuscript, approved the final version and agreed to publish it in this journal.

Acknowledgment

First, the department of Medical Microbiology, School of Biomedical and Laboratory Sciences, College of Medicine and Health Sciences, and University of Gondar are highly acknowledged for facilitating the conduct and data collection in the region. The Amhara Public Health Institute is also acknowledged for allowing molecular detection and further storage of samples. The Armauer Hansen Research Institute and the Ethiopian Public Health Institute are acknowledged for sequencing the positive samples. Finally, the authors would like to thank the health facility leaders, data collectors, and study participants, as their involvement in this study was crucial.

Corresponding author: ORCID: Dessie Tegegne: 0000-0003-3469-017X

Reference

- Burke RM, Hall AJ. Global Burden of Norovirus. *Norovirus*. 2019:1-29.
- Iturriza-Gómara M, Cunliffe NA. *Viral gastroenteritis. Hunter's Tropical Medicine and Emerging Infectious Diseases*: Elsevier; 2020. p. 289-307.
- Naghavi M, Abajobir AA, Abbafati C, Abbas KM, Abd-Allah F, Abera SF, et al. Global, regional, and national age-sex specific mortality for 264 causes of death, 1980–2016: a systematic analysis for the Global Burden of Disease Study 2016. *The lancet*. 2017;390(10100):1151-210.
- Gessese GT, Beksisa J, Mohammed S, Bitew S, Berheto TM, Agachew M, et al. Diarrheal Disease Incidence and Associated Mortality in Ethiopia: A Systematic Subnational Analysis in Global Burden of Disease Study, 1990–2019. *Ethiopian Journal of Health Development*. 2023;37(2).
- Alebel A, Tesema C, Temesgen B, Gebrie A, Petrucka P, Kibret GD. Prevalence and determinants of diarrhea among under-five children in Ethiopia: a systematic review and meta-analysis. *PloS one*. 2018;13(6):e0199684.
- Birhan TA, Bitew BD, Dagne H, Amare DE, Azanaw J, Genet M, et al. Prevalence of diarrheal disease and associated factors among under-five children in flood-prone settlements of Northwest Ethiopia: A cross-sectional community-based study. *Frontiers in Pediatrics*. 2023;11:1056129.
- Bányai K, Estes MK, Martella V, Parashar UD. *Viral gastroenteritis*. *Lancet*. 2018;392:175-86.
- Ahmed SM, Hall AJ, Robinson AE, Verhoef L, Premkumar P, Parashar UD, et al. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and meta-analysis. *The Lancet infectious diseases*. 2014;14(8):725-30.
- Afewerk DT, Shumie MK, Endalew GF, Adugna AG, Tarekegn BG. Pooled prevalence and genetic diversity of norovirus in Africa: a systematic review and meta-analysis. *Virology Journal*. 2022;19(1):1-13.
- Bartsch SM, Lopman BA, Ozawa S, Hall AJ, Lee BY. Global economic burden of norovirus gastroenteritis. *PloS one*. 2016;11(4):e0151219.
- Nordgren J, Svensson L. Genetic Susceptibility to Human Norovirus Infection: An Update. *Viruses*. 2019;11(3).
- Chhabra P, de Graaf M, Parra GI, Chan MC-W, Green K, Martella V, et al. Updated classification of norovirus

- genogroups and genotypes. *Journal of General Virology*. 2019;100(10):1393-406.
13. Farahmand M, Moghoofei M, Dorost A, Shoja Z, Ghorbani S, Kiani SJ, et al. Global prevalence and genotype distribution of norovirus infection in children with gastroenteritis: a meta-analysis on 6 years of research from 2015 to 2020. *Reviews in medical virology*. 2022;32(1):e2237.
 14. Fang Y, Zhang Y, Wang H, Shi O, Wang W, Hou M, et al. Molecular epidemiology of norovirus infections in children with acute gastroenteritis in 2017–2019 in Tianjin, China. *Journal of Medical Virology*. 2022;94(2):616-24.
 15. Udompat P, Srimuang K, Doungngern P, Thippamom N, Petcharat S, Rattanatumhi K, et al. An unusual diarrheal outbreak in the community in Eastern Thailand caused by Norovirus GII. 3 [P25]. *Virology Journal*. 2024;21(1):21.
 16. Chen L, Xu D, Wu X, Liu G, Ji L. An increasing prevalence of non-GII. 4 norovirus genotypes in acute gastroenteritis outbreaks in Huzhou, China, 2014-2018. *Archives of Virology*. 2020;165(5):1121-8.
 17. Wei N, Ge J, Tan C, Song Y, Wang S, Bao M, et al. Epidemiology and evolution of Norovirus in China. *Human Vaccines & Immunotherapeutics*. 2021;17(11):4553-66.
 18. Teunis P, Sukhrie F, Vennema H, Bogerman J, Beersma M, Koopmans M. Shedding of norovirus in symptomatic and asymptomatic infections. *Epidemiology & Infection*. 2015;143(8):1710-7.
 19. Ge Y, Billings WZ, Opekun A, Estes M, Graham D, Leon J, et al. Effect of norovirus inoculum dose on virus kinetics, shedding, and symptoms. *Emerging infectious diseases*. 2023;29(7):1349.
 20. De Graaf M, van Beek J, Koopmans MP. Human norovirus transmission and evolution in a changing world. *Nature Reviews Microbiology*. 2016;14(7):421-33.
 21. Tsang TK, Chen T-M, Longini Jr IM, Halloran ME, Wu Y, Yang Y. Transmissibility of norovirus in urban versus rural households in a large community outbreak in China. *Epidemiology*. 2018;29(5):675-83.
 22. Nordgren J, Svensson L. Genetic susceptibility to human norovirus infection: an update. *Viruses*. 2019;11(3):226.
 23. Lindesmith LC, Brewer-Jensen PD, Mallory ML, Jensen K, Yount BL, Costantini V, et al. Virus–Host Interactions Between Nonsecretors and Human Norovirus. *Cellular and Molecular Gastroenterology and Hepatology*. 2020;10(2):245.
 24. Liao Y, Xue L, Gao J, Wu A, Kou X. ABO blood group-associated susceptibility to norovirus infection: a systematic review and meta-analysis. *Infection, Genetics and evolution*. 2020;81:104245.
 25. Hutson AM, Atmar RL, Marcus DM, Estes MK. Norwalk virus-like particle hemagglutination by binding to H histo-blood group antigens. *Journal of virology*. 2003;77(1):405-15.
 26. Cooling L. Blood groups in infection and host susceptibility. *Clinical microbiology reviews*. 2015;28(3):801-70.
 27. Hutson AM, Atmar RL, Graham DY, Estes MK. Norwalk virus infection and disease is associated with ABO histo-blood group type. *The Journal of infectious diseases*. 2002;185(9):1335-7.
 28. Rockx BH, Vennema H, Hoebe CJ, Duizer E, Koopmans MP. Association of Histo–Blood Group Antigens and Susceptibility to Norovirus Infections. *The Journal of Infectious Diseases*. 2005;191:749-54.
 29. Nordgren J, Kindberg E, Lindgren P-E, Matussek A, Svensson L. Norovirus gastroenteritis outbreak with a secretor-independent susceptibility pattern, Sweden. *Emerging infectious diseases*. 2010;16(1):81.
 30. Nordgren J, Nitiema LW, Ouermi D, Simpore J, Svensson L. Host genetic factors affect susceptibility to norovirus infections in Burkina Faso. *PloS one*. 2013;8(7):e69557.
 31. Bucardo F, Kindberg E, Paniagua M, Vildevall M, Svensson L. Genetic susceptibility to symptomatic norovirus infection in Nicaragua. *Journal of medical virology*. 2009;81(4):728-35.
 32. Liu J, Zhang S, Wang Q, Shen H, Zhang Y, Liu M. Frequencies and ethnic distribution of ABO and RhD blood groups in China: a population-based cross-sectional study. *BMJ open*. 2017;7(12):e018476.
 33. Ahmed M, Memon A, Iqbal K. Distribution pattern of ABO and Rhesus blood groups among different ethnic population of Karachi. *J Pak Med Assoc*. 2019;69(10):1474.
 34. Bruggink L. The epidemiology and evolution of norovirus in Australia. 2016.
 35. Tohma K, Lepore CJ, Ford-Siltz LA, Parra GI. Phylogenetic analyses suggest that factors other than the capsid protein play a role in the epidemic potential of GII. 2 norovirus. *MSphere*. 2017;2(3):10.1128/mspheredirect.00187-17.

36. Sarmiento SK, de Andrade JdSR, Miagostovich MP, Fumian TM. Virological and Epidemiological Features of Norovirus Infections in Brazil, 2017–2018. *Viruses*. 2021;13(9):1724.
37. Almand EA, Moore MD, Jaykus L-A. Norovirus binding to ligands beyond histo-blood group antigens. *Frontiers in microbiology*. 2017;8:2549.
38. Yuan L, Hensley C, Mahsoub HM, Ramesh AK, Zhou P. Microbiota in viral infection and disease in humans and farm animals. *Progress in molecular biology and translational science*. 2020;171:15-60.
39. Gelaw A, Pietsch C, Mann P, Liebert U. Molecular detection and characterisation of sapoviruses and noroviruses in outpatient children with diarrhoea in Northwest Ethiopia. *Epidemiology & Infection*. 2019;147.
40. Dessie TegegneID1, Aschalew Gelaw1, DHA, TS, Leta4 D, GF, et al. Genetic diversity and distribution of noroviruses among all age groups of patients with diarrhea in Amhara National Regional State, Ethiopia. *PloS one*. 2024;19(5).
41. Mujahid A, Dickert FL. Blood group typing: from classical strategies to the application of synthetic antibodies generated by molecular imprinting. *Sensors*. 2015;16(1):51.
42. Yamamoto F, Cid E, Yamamoto M, Blancher A. ABO research in the modern era of genomics. *Transfusion medicine reviews*. 2012;26(2):103-18.
43. Zhuang Z-l, Jin Y, Yan K-l, Cheng W-x. Study of the association between histo-blood group antigens and norovirus infection in Chinese children. *Archives of virology*. 2017;162(11):3511-5.
44. Peña-Gil N, Santiso-Bellón C, Gozalbo-Rovira R, Buesa J, Monedero V, Rodríguez-Díaz J. The role of host glyco-biology and gut microbiota in rotavirus and norovirus infection, an update. *International journal of molecular sciences*. 2021;22(24):13473.
45. Someya Y. Lewis b antigen is a common ligand for genogroup I norovirus strains. *FEBS Open bio*. 2022;12(9):1688-95.
46. Tarris G, Estienney M, Daval-Frérôt P, Lariotte A-C, Aubignat D, Sé K, et al. Intestinal norovirus binding patterns in nonsecretor individuals. *Journal of virology*. 2022;96(19):e00865-22.
47. Cao R-R, Ma X-Z, Li W-Y, Wang B-N, Yang Y, Wang H -R, et al. Epidemiology of norovirus gastroenteritis in hospitalized children under five years old in western China, 2015–2019. *Journal of Microbiology, Immunology and Infection*. 2021;54(5):918-25.
48. Kumazaki M, Usuku S. Norovirus genotype distribution in outbreaks of acute gastroenteritis among children and older people: an 8-year study. *BMC Infectious Diseases*. 2016;16:1-8.
49. Lian Y, Wu S, Luo L, Lv B, Liao Q, Li Z, et al. Epidemiology of norovirus outbreaks reported to the public health emergency event surveillance system, China, 2014–2017. *Viruses*. 2019;11(4):342.
50. Simon AK, Hollander GA, McMichael A. Evolution of the immune system in humans from infancy to old age. *Proceedings of the Royal Society B: Biological Sciences*. 2015;282(1821):20143085.
51. Shamkhali Chenar S, Deng Z. Environmental indicators for human norovirus outbreaks. *International journal of environmental health research*. 2017;27(1):40-51.
52. Chiu S-C, Hu S-C, Liao L-M, Chen Y-H, Lin J-H. Norovirus Genogroup II Epidemics and the Potential Effect of Climate Change on Norovirus Transmission in Taiwan. *Viruses*. 2022;14(3):641.